

Microsampling of Non-Invasive Matrices: Practical Examples Using Tears and a Perspective of Past and Emerging Technologies

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Jason Watts, PhD



Session Description and Objectives

Non-invasive matrices such as tears, sweat, saliva, and milk offer an enticing alternative to traditional sampling from blood, serum, and plasma. Collection of these matrices is simpler and more affordable than venipuncture, as well as being much less unpleasant for patients and study subjects allowing for more frequent sampling, higher compliance, and removal of fear of needles as a barrier to clinical trial recruitment. However, bioanalysis of these matrices is not without challenges. Sample volumes can be very low, the more complex matrices such as milk may require extensive sample preparation, and the use of the matrix must be biologically relevant. Using the example of a method we developed at Alturas Analytics Inc. to measure Tobramycin from human tears, we will discuss practical applications of microsampling of non-invasive matrices for bioanalysis, and improvements offered by emerging technologies.

- Understand the benefits and challenges of using non-invasive matrices in bioanalysis
- Explore currently available microsampling technologies and advances in analytical techniques
- Discuss strategies for collecting, extracting, and analyzing non-invasive matrices



Microsampling

- Typically uses $\leq 50 \mu\text{L}$
- Simple collection and storage
- Possibility for at-home sampling
- Site-Centric \longrightarrow Patient-Centric



Non-invasive matrices

- Tears, sweat, saliva, milk
- High correlation with non-protein bound plasma concentrations for many drugs



Practical considerations

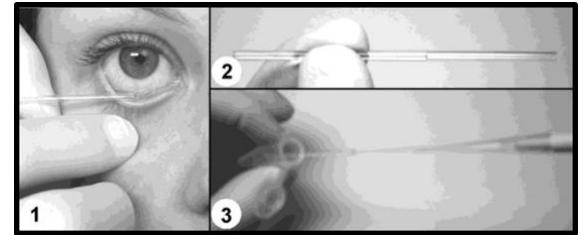
- Can sufficient sensitivity be achieved?
- Analyte stability
- Does the matrix concentration correlate with plasma concentrations?
- What is gained over traditional sampling?



Tears

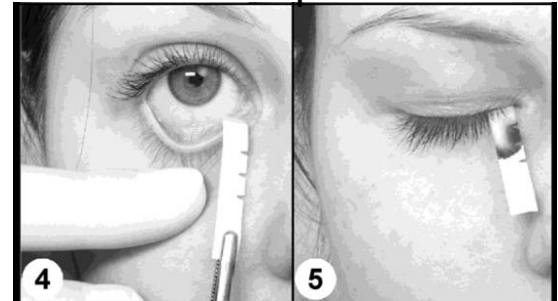
- Constantly produced at 1.2 $\mu\text{L}/\text{min}$
- Unstimulated volume $\sim 7 - 10 \mu\text{L}$
- 0.6 - 0.8% protein (0.4% albumin)
- pH 6.5 - 7.6
- Differences between stimulated and unstimulated collection

Capillary tube collection



From Posa et al., 2012

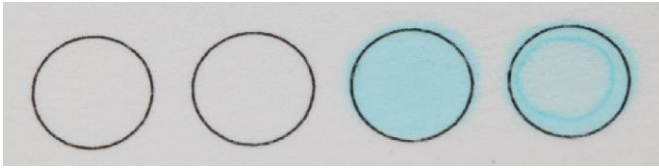
Schirmer strip collection



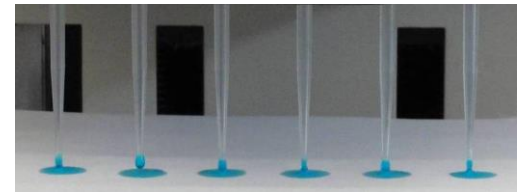
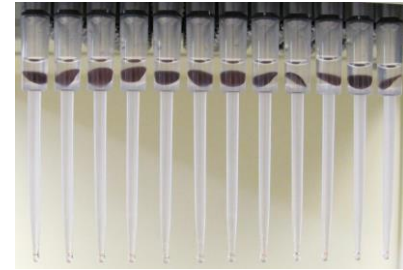
From Posa et al., 2012



Adapting blood microsampling techniques for non-invasive matrices



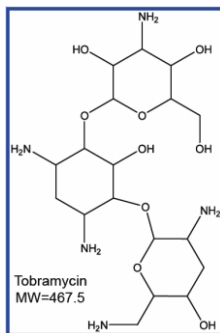
Improved precision and accuracy using color-indicating dyes



Improving extraction workflow with common Bioanalytical tools



Tobramycin from human tears



QC Level ($\mu\text{g/mL}$)	Assay Accuracy and Precision (% \pm %CV)	Matrix Factor
12	90.0 \pm 4.5	NA
3.0	96.2 \pm 0.07	NA
1.5	101 \pm 5.7	0.97



Sweat

- Collected on sweat wipes or patches
- Low sample volume/need for normalization
- pH 4 - 6.8 when resting.
 - High sweat/plasma ratio for basic drugs
- Commonly used for monitoring drugs of abuse



Saliva

- Production and composition
 - 0.5mL/min
 - pH ~ 6 - 7
 - Excretion of drugs dependent on permeability and protein binding
- Noninvasive collection
 - Unstimulated
 - Stimulated



Fig. 4. A selection of different swab-based devices for sampling saliva: (A) Super Sal™ (Oasis Diagnostic Corporation); (B) Versi Sal™ (Oasis Diagnostic Corporation); (C) Quantalife™ (Immunalysis Corporation); (D) Intercept® (Orasure Technologies Inc.); (E) Salivette® (blue cap, Sarstedt); (F) S05 (Salimetrics); (G) Toothette-Plus swabs (Sage Products Inc.); (H) OraQuick Advance HIV-1/2 (Orasure Technologies Inc.); (I) BBL CultureSwab orange and white cap.

Bellagambi et al., 2020



Using blood microsampling device for saliva sampling

Mitra[®] (Neoteryx) microsampling device

- Precise sample collection
- Ideal for pediatric or at home sampling
- 10, 20, or 30 μL sample volume



Correlation between plasma and saliva dexamethasone concentrations

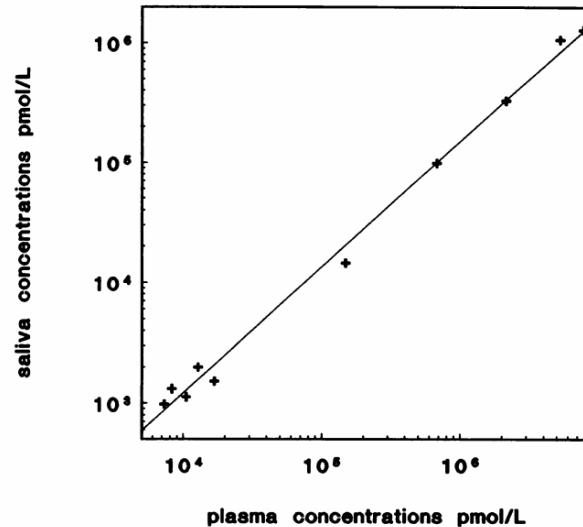


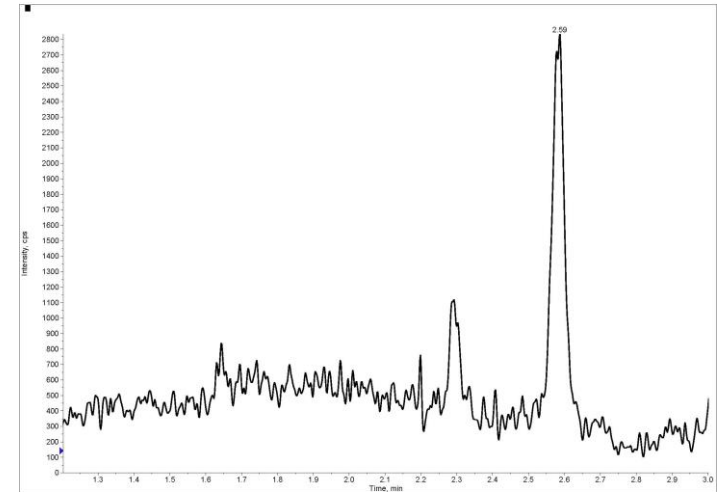
Fig. 2. Correlation between concentrations of dexamethasone in plasma and saliva taken simultaneously from healthy subjects and patients pretreated with dexamethasone.

Thijssen et al., 1996



Simple extraction of dexamethasone from saliva using a Mitra[®] Microsampling device

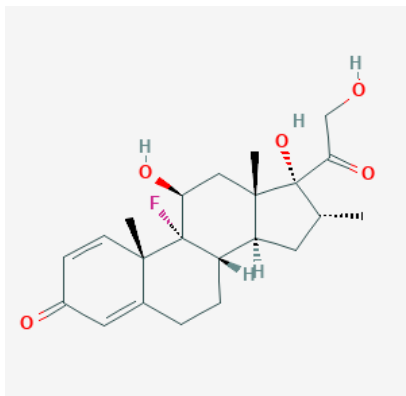
1. Collect sample on 20 μ L Mitra[®] tip
2. Dry for at least 2 hours at ambient temperature
3. Place dried tip in 96 well plate with curve and QCs
4. Add 25 μ L 100 ng/mL IS (Dex-D4 in ACN:H₂O 1:1)
5. Add 300 μ L ACN. Vortex and incubate 30 min
6. Transfer extract to fresh plate. Evaporate to dryness
7. Reconstitute in 100 μ L of ACN:H₂O with 0.1% formic acid



0.4 ng/mL Dexamethasone extracted from Saliva



Dexamethasone extracted from saliva



Dexamethasone 392.5 g/mol

Concentration (ng/mL)	Assay Accuracy and Precision (% \pm % CV)	% Recovery
0.8	98.3 \pm 0.94%	94
50	103.5 \pm 0.68%	117
400	100.1 \pm 7.0%	121

Dexamethasone structure from <https://pubchem.ncbi.nlm.nih.gov/compound/Dexamethasone>



References

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- Andreas Posa, Lars Brauer, Martin Schicht, Fabian Garreis, Stephanie Beileke, and Friedrich Paulsen., Schirmer strip vs. capillary tube method: Non-invasive methods of obtaining proteins from tear fluid. Annals of Anatomy 195 137-142. 2013
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Jennizer Zimmer

Chad Christianson

Cody Hawkins



Questions and Contact Information

AlturasAnalytics.com
info@alturasanalytics.com
(208)883-3400

1324 Alturas Dr.
Moscow, ID 83843

